Characterization of dynamics and internal structure of a mixed-surfactant wormlike micellar system using NMR and rheometry

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We use complementary experiments—proton NMR diffusometry and relaxometry, deuterium NMR lineshapes, and rheometry—to construct a comprehensive picture of the microscopic structure of a mixed-surfactant wormlike micellar system composed of a zwitterionic surfactant and an anionic surfactant in brine. In this system, at some surfactant concentrations, the time for micellar breaking and recombination \( t_b \) is not small compared with the micellar reptation time \( \tau_R \), weakening the condition to obtain a stress relaxation function with just one relaxation time at long times. From NMR relaxometry, we determine the overlap concentration. Deuterium NMR spectral lineshapes indicate the presence of a wide angular distribution in the orientational order. NMR diffusometry and rheology probe different timescales and yield complementary information indicating polymer-like behaviour at the corresponding length scales. Via NMR, surfactant diffusion coefficients are seen to decrease with increasing diffusion time, consistent with restricted diffusion within a reptating micelle. At the same time, comparison of measurements with protonated and deuterated surfactants strongly suggests that the measured short and long time diffusion coefficients correspond to intra-micellar and micellar diffusion, respectively. Fitting the diffusion results to a simple model, the average end-to-end micellar distance is estimated to be in the 1 \( \mu \)m range and only weakly dependent on concentration. The water diffusion measurements, on the other hand, imply a high degree of water structuring at the micellar surface. We also find that the wormlike micelles obeyed simple polymer-like scaling behaviors, with a crossover from Zimm-like (diffusion) to Rouse-like (rheology) exponents.

Introduction

Amphiphilic molecules in aqueous solution form aggregates with different structures such as bilayers, vesicles, and spherical, cylindrical or wormlike micelles, depending on molecular geometry as well as on the net charge and surfactant concentration.1 Wormlike micelles are interesting, due to the fact that they are elongated objects like polymers (which have interesting dynamics and hydrodynamic effects2–5); however, they continuously break and recombine.6–10 This has technological applications (from heat-transfer fluids to oil-field applications to drain-openers)9 because, unlike normal polymers, they can reform after breaking and can thus survive repeated shear.6

Much work has been carried out on the study of wormlike micelles via theory6,11–13 and computer simulation,14,15 and there have been several reviews on the subject.6,7,10,15,16 Experimentally, wormlike micellar systems can be composed of cationic, anionic, zwitterionic or non-ionic surfactants.6,8,17–20 Mixtures of cationic and anionic surfactants,20 zwitterionic dimeric surfactant solutions21 and cationic surfactant solutions20,22 have also been investigated. A common surfactant in wormlike micelle literature is cetyl trimethylammonium bromide (CTAB)23–27 which forms cylindrical wormlike surfactant micelles; systems forming reverse micelles have also been reported.28,29

Small-angle neutron scattering and dynamic light scattering studies have reported on isotropic to nematic phase transitions,23 transitions from vesicles to wormlike micelles,30 the concentration dependence of the hydrodynamic correlation length,31 and the effect of adding salt on micellar growth.32 Scattering measurements have also characterized important lengths of the micellar network,33 and have been used to investigate local structure and flexibility.34,35 A combination of rheology and small angle neutron scattering (Rheo–SANS) has also been used to study concentration dependencies of shear thinning and alignment in a block copolymer wormlike micellar system.36

Pulsed-field-gradient nuclear magnetic resonance was used to identify sub-diffusive behavior in a wormlike reverse micelle system.37 This technique has been employed in other (polymer and protein) soft matter systems38,39 to provide information that is complementary to scattering methods, and is especially useful when the system contains large and/or multi-component aggregates. This is because spectral separation of different chemical
components is easy in NMR, challenging in scattering, and practically impossible in rheology.

Recently, a multi-component system consisting of a mixture of two similar-sized surfactants, one zwitterionic (N-tetradecyl-N,N-dimethyl-3-ammonio-1-propanesulfonate or TDPS) and the other anionic (sodium dodecyl sulfate or SDS), in brine\textsuperscript{33,40,41} has been studied by rheology and scattering techniques. The TDPS–SDS system was studied at a range of TDPS concentrations \(C_z\), spanning both dilute and semidilute regimes at different surfactant ratios \(R = [SDS]/[TDPS]\) and different temperatures.\textsuperscript{33,40,41} The average micelle contour length was found to be in the micron range and (for \(R = 0.55\)) to increase with \(C_z\) when the wormlike micelles are totally screened by salt addition, while the “mesh size” reflecting intermicellar correlations is in the 50–100 nm regime and insensitive to changes in \(C_z\), surfactant concentration ratio or temperature.\textsuperscript{33} An interesting feature of this system is that while \(R = 0.55\) mixtures could be fitted to Maxwellian viscoelastic behavior with a single relaxation time for the stress relaxation modulus \(G(t)\) at long times,\textsuperscript{41} those for \(R = 0.43–0.45\) show deviations from the Maxwellian model.\textsuperscript{41} Two timescales are relevant in wormlike micellar systems. \(\tau_0\) refers to the breaking/recombination timescale, while \(\tau_R\) is the reptation timescale. If the micellar chain breaks and recombines like contour segments exhibit a single relaxation rate.\textsuperscript{8}

In our previous work,\textsuperscript{38} we have shown that mixed-species (polymer–surfactant) aggregates are easier to study than single-species (surfactant) aggregates because the dynamics of each species can be independently and simultaneously measured via complementary NMR experiments. This allows models of aggregate structure to be sufficiently constrained. In this work, using the TDPS–SDS system (with \(R = 0.45\)), we obtain in unprecedented detail both the structure and dynamics in a system where the linear rheology is not dominated by a single relaxation time at low frequencies.

In this work, we use NMR diffusometry and relaxation measurements, as well as deuterium NMR, to explore the dynamics and the structure of the micellar aggregates. Relaxation in NMR usually refers to two processes by which nuclear magnetization prepared in a non-equilibrium state returns to the equilibrium distribution. Different physical processes are responsible for the relaxation of the components of the nuclear spin magnetization vector \(M\), parallel and perpendicular to the external magnetic field, \(B_0\), which is conventionally oriented along the z-axis. These two principal relaxation processes are termed \(T_1\) and \(T_2\) relaxation, respectively. The longitudinal \(T_1\) and transverse \(T_2\) relaxation times can be measured directly using NMR, and can be used to report on changes in the local environment.\textsuperscript{42}

In the NMR diffusion experiment, the sample experiences both an external uniform magnetic field from the magnet and a non-uniform spatially well-defined magnetic field (i.e. pulsed field gradient). Therefore, the molecular diffusion is measured from the signal attenuation that arises from the dephasing of nuclear spin coherence.\textsuperscript{43,44} Deuterium NMR, on the other hand, is an effective probe of the orientational order of the deuterated hydrocarbon chains.\textsuperscript{44} Thus, we are able to obtain independent dynamical information on all components in a 3-component system via spectral separation (via a difference in either chemical shift or spin label) of diffusion coefficients.

Utilizing these complementary NMR techniques on the TDPS–SDS system (with \(R = 0.45\)), in tandem with rheology, we obtain in unprecedented detail both the structure and dynamics in the case where the linear rheology of the system is not dominated by a single relaxation time at low frequencies.

**Experimental**

**Materials**

\(N\)-Tetradecyl-N,N-dimethyl-3-ammonio-1-propanesulfonate (TDPS, \(M_w = 363.6\), purity >99%), sodium dodecyl sulfate (SDS, \(M_w = 288.38\), purity >99%), and SDS-d25 (\(M_w = 313.53\), 98 atom% D) were purchased from Sigma-Aldrich Canada and were used as received. We prepared stock solutions of TDPS–SDS(0.5 M)–D\(_2\)O as well as TDPS–SDS-d25. NaCl(0.5 M)–H\(_2\)O at \(C_z = 50\) mM, as well as the brines NaCl(0.5 M)–D\(_2\)O and NaCl(0.5 M)–H\(_2\)O. Samples with TDPS concentration and protonated/deuterated SDS below 50 mM (in the semidilute regime) were prepared by diluting with brine made with D\(_2\)O/H\(_2\)O respectively. The sample in the concentrated regime \(C_z = 140\) mM was prepared separately. The surfactant ratio in all samples was \(R = [SDS]/[TDPS] = 0.45\). Deuterium oxide D\(_2\)O (for all protonated SDS samples) and deuterium-depleted H\(_2\)O (for deuterated SDS samples) were purchased from Cambridge Isotope Laboratories. While surfactant mixtures were prepared by addition of constituents by mass, the volume fraction is the relevant quantity when considering hydrodynamic corrections.

We estimate a volume fraction \(\Phi\) using

\[
\Phi = \frac{V_{TDPS} + V_{SDS}}{V_{TDPS} + V_{SDS} + V_{NaCl(0.5M)}-H_2O} 
\]

where \(V_a = m_a/\rho_a\), \(V_a, m_a\) and \(\rho_a\) are the volume, mass and density of the \(a\) component in solution, and \(\alpha\) indicates TDPS, SDS or NaCl(0.5 M)–H\(_2\)O. Note that this is an estimate of volume fraction due to the assumption of volume additivity. Because of the densities (\(\rho_{SDS} = 1.09\) g cm\(^{-3}\), \(\rho_{NaCl(0.5M)}-H_2O = 1.02\) g cm\(^{-3}\)), our estimated volume fraction is essentially equivalent to the mass fraction.

NMR spectra were collected on a Bruker Avance II spectrometer with a \(^1\)H resonance frequency of 600.33 MHz and a \(^2\)H resonance frequency of 92.15 MHz. We can easily separate the water peak at 4.7 ppm and the surfactant peaks spectrally in the NMR spectrum. However, due to the broad (super-Lorentzian) lineshapes, we are unable to spectrally separate the TDPS and SDS peaks (0–4 ppm). Thus, we prepared some samples with deuterated SDS. We were then able to compare the surfactant dynamics of TDPS (proton NMR with deuterated SDS yields only the TDPS peaks) with the surfactant dynamics of TDPS-SDS peaks in protonated SDS samples.

Relaxation measurements were performed using a Micro-5 imaging (3-axis gradient) probe. The inversion recovery technique was used to measure \(T_1\): sixteen time delays were used, and the integrated intensities in the spectrum were fitted to the equation: \(I(t) = I_0(1 - 2\exp(-t/T_1))\).\textsuperscript{42} A \((\pi/2)_y - t/2 - (\pi)_z - t/2\)-acquire spin echo experiment was used to measure \(T_2\): 16 values, at delay time \(t\), of the integrated intensity were taken to measure \(T_2\) and were fitted to the equation: \(I(t) = I_0 \exp(-t/T_2)\).\textsuperscript{42}
Three-axis self-diffusion measurements were carried out in a Micro-5 imaging (3-axis gradient) probe with a maximum gradient strength of 200 G cm$^{-1}$ or a Diff30 diffusion probe with a maximum field gradient 1800 G cm$^{-1}$, employing a pulsed-field gradient stimulated-echo sequence$^{44}$ with (almost square) trapezoidal gradient pulses, with the rise time of the trapezoid (the gradient ramp time) being 100 $\mu$s, i.e. much smaller than the diffusion time $\Delta$ or the total duration $\delta$ of the field gradient pulse. In our experiments, $\delta = 2$ ms, and the gradient $g$ was varied in steps from 0% to $\approx 100\%$. For deuterated SDS samples, $^2$H NMR spectra were collected using a $(\pi/2)_x$–$t$–$(\pi/2)_y$–$t$-acquire quadrupole echo experiment using 256 scans, and with $t = 10 \mu$s.

Spectrally resolved molecular diffusion coefficients along the field direction ($D_z$) and perpendicular to the field direction ($D_x$ and $D_y$) are obtained from the attenuation of the signal according to the equation:$^{45}$

$$D = \frac{-1}{k} \ln \left( \frac{S(k)}{S(0)} \right)$$

(2)

where $S(k)$ is the integrated intensity of the signal in the presence of the field gradient, while $S(0)$ is the intensity of the signal in the absence of the field gradient, and $k = (\gamma g/\delta)(\Delta - \delta/3)$ is a generalized gradient strength parameter. The diffusion time $\Delta$ is the duration of the pulse sequence in which molecular diffusion has an effect on the signal attenuation. The signal attenuation as a function of $k$ associated with TDPS–SDS peaks is a single exponential (Fig. 1) over the whole range of TDPS concentrations. When there is free bulk diffusion, the diffusion coefficient does not depend on the diffusion time $\Delta$.

**Fig. 1** Natural logarithm of the signal attenuation *versus* the gradient strength parameter $k = (\gamma g/\delta)(\Delta - \delta/3)$ with $\Delta = 100$ ms for (top) TDPS(30 mM)–SDS(13.5 mM)–NaCl(0.5 M)–D$_2$O and (bottom) TDPS(50 mM)–SDS(22.5 mM)–NaCl(0.5 M)–D$_2$O samples at $T = 298$ K. The spectral region 0–4 ppm is used to measure the surfactant signal attenuation.

**Rheometry**

Rheological measurements were carried out on an Anton Paar Physica MCR 301 rheometer. All the rheometric measurements were done at $T = 298$ K using the cone–plate geometry of 50 mm diameter and 0.5° cone angle. The stress relaxation experiments were performed with an applied shear strain $\gamma = 0.5$. The flow curve experiments were carried out with shear strain rate $\dot{\gamma}$ varying from 0.001 to 150 s$^{-1}$ to extract the zero-shear strain viscosity $\eta$. In addition, the oscillatory shear experiments were performed with an angular frequency $\omega$ varied in log-ramp from 50 rad s$^{-1}$ to 0.01 rad s$^{-1}$.

**Results and discussion**

**NMR relaxation and the overlap concentration**

Longitudinal $T_1$ and transverse $T_2$ relaxation measurements were carried out for TDPS–SDS–NaCl(0.5 M)–D$_2$O samples at different TDPS concentrations, $C_z$. The proton longitudinal relaxation time $T_1$ (Fig. 2) can be fit to a mono-exponential decay, with a significant decrease as one crosses the overlap concentration (reported to be at 7 mM by Lopez-Diaz *et al.*$^{40}$). The change in $T_1$ indicates a change in the local environment for the surfactant molecules, which corresponds well with the wormlike micelle overlap concentration. Based on the exponential fit, we extract a characteristic concentration of $C_{\text{threshold}} = 4.5 \pm 0.4$ mM (*i.e.* $C_{\text{threshold}} \approx C^*$. The $T_2$ relaxation time (not shown) shows no appreciable change as a function of TDPS concentration $C_z$.

**Deuterium NMR and orientational structure**

Measurements with deuterated SDS provide good opportunities to look at the orientational structure of the SDS (using deuterium NMR), and to separate the dynamics of the TDPS (using proton diffusometry). We also use these samples to measure the variation of the water diffusion coefficient with surfactant concentration.

Fig. 3a shows $^2$H NMR spectra for $C_z = 46$ mM as a function of temperature. The appearance of a single broad peak in the deuterium NMR spectrum implies a structure that is intermediate between an isotropic liquid and an oriented liquid. It is likely an indication of the presence of a liquid crystal like mesophase with a wide orientational angular distribution. As we will describe below, this is consistent with the interpretation of

![Fig. 2](image-url) Proton longitudinal relaxation time $T_1$ *versus* TDPS concentration $C_z$ for TDPS–SDS–NaCl(0.5 M)–D$_2$O samples at $T = 298$ K.
Fig. 4a, where we see only a small anisotropy in the measured diffusion coefficient. This averages out the first-order quadrupole coupling.\cite{46} We may fit the deuterium NMR lineshape to the absorption Lorentzian function

\[ S(\omega) = \frac{(W/2)^2 + (\omega - \omega^0)^2}{(W/2)^2 + (\omega - \omega^0)^2} \]  

(3)

where \( \omega \) and \( \omega^0 \) are the frequency coordinates and the Larmor frequency of the deuterium (spin-1) nuclei, respectively. \( W \) is the deuterium peak width at half maximum of the absorption peak.

For deuterated SDS samples (TDPS–SDS–d25–NaCl(0.5 M)–H2O) at two TDPS concentrations (\( C_z \approx 10 \) and 46 mM), we can extract the deuterium peak width at half maximum (W) of the absorption peak as a function of temperature between 298 K and 323 K. The width of the SDS peak at 298 K is \( /C24 200 \text{ Hz} \) (about a factor of 10 larger than an HDO peak when deuterium-depleted water is not used). In general, a single-Lorentzian peak could result either from exchange mediated by a free surfactant monomer in solution or by the surfactant seeing a continuous change in orientational order while diffusing inside a curvilinear micelle. Given the peak width, diffusion inside a micelle with a continuously changing orientation seems to dominate the signal represented by the broad isotropic peak. Using a surfactant diffusion coefficient \( D_s = 13 \times 10^{-12} \text{ m}^2 \text{ s}^{-1} \), and the experimental timescale of 10 µs yields a lengthscale of \( \ell = \sqrt{6\pi D_s \approx 28 \text{ nm}} \). This is roughly consistent with the values (~35 nm) reported for the persistence length in this system.\cite{33} A similar broadening of a broad isotropic peak with decreasing temperatures has been seen in so-called “stiff gel” smectics in the presence of a high density of quenched random disorder, which has been interpreted, not as a true isotropic phase, but as a smectic phase with a wide and smooth angular distribution of the orientational order.\cite{45} This peak width shows an Arrhenius temperature dependence. By globally fitting both data sets in Fig. 3b to one exponential function \( W = A \exp(E_a/k_B T) \), we estimate the activation energy to be \( E_a \approx 21.8(1)k_B T \approx 54 \text{ kJ mole}^{-1} \); this represents the barrier for surfactant diffusion in these wormlike micelles.

Water diffusion in aqueous surfactant solutions has been shown to be a weighted average of two kinds of water, bulk water and surface associated water.\cite{47} The water molecule is polar, and it associates with a surface of charged molecules or charged aggregates. It has been shown to be a reasonable approximation to assume that the surface-associated water is essentially stationary in comparison to the bulk water.\cite{36,47} Thus, water diffusion coefficient obtained as a function of surfactant packing fraction report directly on the fraction of surface associated water. Fig. 3c shows the variation of the relative self-diffusion...
The observed self-diffusion coefficient (Fig. 4a) measured along the z, x, and y axes displays a very similar behaviour, and thus exhibits insignificant ordering effects in the $\equiv 14$ T magnetic field. $D_z$, $D_x$, and $D_y$ all decrease as a function of diffusion time $\Delta$. If what we measure is only the self-diffusion of the wormlike micelle, then we would not expect $D$ to show a dependence on $\Delta$. For $\Delta \approx 15$ ms, the wormlike micelle is already in the long-time limit due to the fact that RMS displacement over the millisecond timescale is much larger than the average mesh size of the TDPS–SDS–NaCl(0.5 M) micellar solution $\xi \approx 75$ nm.$^{31}$ The observed decrease of $D$ with $\Delta$ suggests strongly that the observed diffusion coefficient is a combination of micellar self-diffusion and surfactant self-diffusion inside the micelle.

This is confirmed by comparing surfactant diffusion coefficients for systems with non-deuterated and fully deuterated SDS. This is akin to contrast matching experiments in neutron scattering. The surfactant self-diffusion coefficient $D_z$ was measured for TDPS–SDS–d$_{25}$–NaCl(0.5 M)–H$_2$O samples with deuterated SDS at 3 TDPS concentrations (40 mM, 46 mM, and 50 mM). Here, we measure the TDPS self-diffusion coefficient, whereas in the protonated samples one measures an average value of TDPS and SDS diffusion coefficients. If the two values are equal, they will lie on the 45 degree line. The values obtained for the relative diffusion coefficients in Fig. 4b (left) are only close to being equal at the smallest values (corresponding to the largest $\Delta$ (500 ms)), where $D[H_2O] = 2.23 \times 10^{-9}$ m$^2$ s$^{-1}$ and $D[NaCl] = 1.73 \times 10^{-9}$ m$^2$ s$^{-1}$ are the diffusion of H$_2$O and D$_2$O, respectively, in 0.5 M brine. This is consistent with the notion that the micellar diffusion coefficient should depend only on the micelle size and the solvent viscosity. Therefore, assuming a Stokes–Einstein-like relationship for the micelle in the solvent $D \propto 1/\eta$, the diffusion coefficient relative to the solvent diffusion coefficient should be the same for both systems.

Fig. 4b (right) on the other hand shows the bare self diffusion coefficients $D_z$. Here we see that the values for the deuterated and non-deuterated samples are closest for small $\Delta$ (100 ms) (i.e. for large $D_z$). This is consistent with surfactant diffusion within the micelle—these values should indeed be insensitive to solvent viscosity.

Three-axis diffusion measurements thus show that there is very insignificant anistropy in the wormlike micelle conformations in the presence of a large magnetic field. Moreover, comparison of the measurements in the deuterated and non-deuterated surfactant show a clear trend: micellar diffusion is dominant for large $\Delta$, while intramicellar diffusion is dominant for small $\Delta$.

### Micellar and intramicellar diffusion

From the observed values of anisotropic self-diffusion coefficients (Fig. 4a) at different diffusion times $\Delta$, the mean-square displacement (MSD) values (MSD$_i = 2D_i\Delta$, $i = x,y,z$) are obtained (Fig. 5a). The dependence is clearly very linear; however the straight line does not pass through the origin. Therefore, the diffusion is neither lateral diffusion$^{48}$ nor is it single-file diffusion$^{49}$ for which the MSD should scale as the square root of the diffusion time. The slope in Fig. 5a represents a reasonable estimate for the micellar diffusion (Fig. 5b) at different TDPS concentrations $C_z$. The non-zero intercept, on the other hand, is an indication of faster intra-micellar surfactant diffusion at shorter times.

Fig. 5b shows the variation of the anisotropic micelle diffusion coefficients ($D_{mx}$, $D_{my}$, and $D_{mz}$) of the micelle as a function of TDPS concentration in the semidilute regime (i.e. 10 mM $\leq C_z \leq$ 50 mM) for ($D_{mx}$), ($D_{my}$), and in the 10 mM $\leq C_z \leq$ 140 mM regime for ($D_{mz}$) of TDPS–SDS–NaCl(0.5 M)–D$_2$O. The anisotropic diffusion curves $D_x$ and $D_y$ are offset along the y-axis. The anisotropic self-diffusion coefficients exhibit a power law decrease with respect to $C_z$.

We globally fit the three datasets in Fig. 5b in the semidilute regime to a single power law (“Global fit” in graph). Our experiments yield a power law $D = D_{Zimm}(C_z/C^*)^{-d}$ with the exponent $d = 0.58 \pm 0.03$. The Zimm model for polymer dynamics in a good solvent considers the hydrodynamic interactions between the monomers on the polymer chain and between the monomers and the solvent molecules in the pervaded volume. It would predict an exponent $d = (1 - v)/(3v - 1) = 0.54$ (using $v = 0.588$ for a self-avoiding polymer).$^2$ This exponent is shown for comparison (dashed line labeled “Zimm model” in the graph). We calculate an experimental exponent $v = (1 + d)/(1 + 3d) = 0.58 \pm 0.01$. From the power law fit, we also obtain average Zimm diffusion coefficient $D_{Zimm}$ of the wormlike micelle of $8.3(1) \times 10^{-12}$ m$^2$ s$^{-1}$.

For free diffusion, $D$ is independent of $\Delta$. For bounded diffusion, the apparent diffusion coefficient is smaller than the true diffusion coefficient at long times, when molecules start to feel the effects of the boundaries. The signal attenuation function $S(q)$, which is related to the diffusion coefficient $D$ via eqn (2), is modified due to the effect of boundaries. We utilize a simple restricted diffusion model for surfactant diffusion in a one
Fig. 5 (a) Z-axis mean square displacement MSD$_z$ versus diffusion time $\Delta$. (b) Anisotropic micelle self-diffusion coefficients ((D$_m)_x$,(D$_m)_y$,(D$_m)_z$), extracted from the slopes of the mean-square displacement MSD curves as a function of TDPS concentration (C$_z$) for TDPS–SDS–NaCl(0.5 M)–D$_2$O samples at $T = 298$ K. The interception are not zero but are on the 0.5 $\mu$m scale. (c) The average 2-end-to-end distance $L_z$ of the TDPS–SDS micelle versus TDPS concentration $C_z$, extracted from a fit of the signal attenuations using eqn (5), for TDPS–SDS–NaCl(0.5 M)–D$_2$O samples at $T = 298$ K. The mean $L_z$ is around 1.1 $\mu$m.

as the diffusion time increases, a larger fraction of molecules feel the effects of confinement, with the diffusion coefficient not changing much for $\Delta \geq 300$ ms. The z-axis signal attenuation curves for each diffusion time in Fig. 4a were fitted to eqn (5). From this fit (signal attenuation curves and fit not shown), we extract the average end-to-end distance $L_z$ (Fig. 5c) for the wormlike micelle. The infinite sum in eqn (5) is approximated by an upper limit $N = 1000$. $D_m$ was obtained from the slopes of Fig. 5a for each TDPS concentration $C_z$ (values obtained for $D_m$ are shown in Fig. 5b). $D_n$, the free diffusion of surfactant inside a micellar environment, and $L_z$ are fit parameters. A value of $D_s = 13 \times 10^{-12}$ m$^2$ s$^{-1}$ provided a good fit for all concentrations. Using the Zimm diffusion coefficient $D_{Zimm}$, the average time $\tau_{Zimm}$ for a micellar chain to diffuse a distance of order of its average end-to-end distance $L$ can be calculated.\(^\dagger\) Using $L_{Zimm} = 1.1(1)$ $\mu$m and $D_z = 8.3(1) \times 10^{-12}$ m$^2$ s$^{-1}$, we get $\tau_{Zimm} = (L_{Zimm})^2/D_{Zimm} \approx 0.15$ s.

Rheology and supramicellar structure

Relaxation modulus experiments were performed for a range of TDPS concentrations in the semidilute regime (10 mM $\leq C_z \leq 50$ mM). If the relaxation modulus exhibits a single exponential at long times, then the recombination and the scission process is rapid, i.e. the micellar chain breaks and recombines many times on a time scale $\tau_b \ll \tau_R$ and the primary relaxation mechanism is reptation.\(^\dagger\)

A stress relaxation dominated by a single exponential relaxation decay is generally consistent with the wormlike character of the self-assembled structures in the solution. If $\tau_R \gg \tau_b$, stress relaxation follows a single exponential in wormlike solutions according to the Cates model. Lopez-Diaz et al.\(^\ddagger\) note that $\tau_b/\tau_R$ is at a minimum at $R \approx 0.55$, which is precisely the $R$ value where the mixture perfectly fits the Maxwellian Cole–Cole semicircle. While stress relaxation results for $R \approx 0.55$ mixtures could be fitted to Maxwellian viscoelastic behavior with a single relaxation time for the stress relaxation modulus $G(t)$,\(^\ddagger\) those for $R \approx 0.43$ showed deviations from the Maxwellian model.\(^\ddagger\) For our $R = 0.45$ samples, all the relaxation functions $G(t)$ (not shown) are better fitted to a bi-exponential than a single exponential.

The elastic modulus $G_0$ (Fig. 6a), associated with the slower, dominant, relaxation mode, is extracted. Reduced viscoelastic spectra are extracted over the whole range of TDPS concentrations $C_z$. The inset in Fig. 6a is an example of a Cole–Cole diagram for $C_z = 50$ mM. The upturn in $G''$ at higher frequencies is an expected outcome of Rouse-like behavior.\(^\ddagger\) This elastic modulus (Fig. 6a) scales as $C_z^{0.55}$ where the exponent $b$ is given by $3v/(3v - 1) = 3.0 \pm 0.1$, implying $v = 0.50 \pm 0.01$. In addition, the zero-shear-strain viscosity (Fig. 6b) also scales\(^\ddagger\) as $C_z^{0.47}$ with $s = 1/(3v - 1) = 2.4 \pm 0.1$, or $v = 0.47 \pm 0.01$. Both these scaling behaviors are in the semidilute regime, and consistent with $(v = 1/2)$ Rouse-like behavior.

Two polymer-like scalings

In this work, we observed two distinct polymer-like scalings in micellar dynamics in the semi-dilute regime. Diffusometry in the semi-dilute regime is consistent with a scaling exponent $v = 0.58 \pm 0.01$ (consistent with the value of 0.588 for Zimm-like behavior in a good solvent). On the other hand, rheological measurements are consistent with a random-walk scaling exponent of $v = 0.5$.

These results have a simple interpretation. In the semidilute regime of mixed-surfactants TDPS–SDS, NMR diffusometry

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the water molecules H$_2$O with respect to the total surfactant functional dependence of the relative self diffusion coefficient of molecule in deuterated SDS samples, we were able to obtain the regime are consistent with the exponent expected for Rouse-like behavior: see text.

The water molecules H$_2$O provide access to dynamics at shorter lengthscales and times (where hydrodynamic interactions are not screened) and can be described by Zimm dynamics in a good solvent. On the other hand, the rheological measurements provide access to dynamics at larger scales at which the hydrodynamic interactions are screened, and Rouse dynamics results. In spite of the fact that the range of the fit is small in order to remain in the semi-dilute regime, the two exponents are distinct (about 8 apart) and are consistent with classic polymer-like scaling in different hydrodynamic regimes.

**Conclusions**

In this work, we carried out several complementary NMR experiments, as well as rheology, and these results yield a comprehensive picture of the microscopic structure in a wormlike micellar system. From NMR longitudinal relaxation $T_1$ measurements (Fig. 2), we estimate the TDPS characteristic overlap concentration $C^* \approx 4.5 \pm 0.4$ mM. This compares reasonably with the value of $\approx 6$–7 mM determined using dynamic light scattering. In addition, the temperature dependent $^2$H NMR spectra (Fig. 3a) show a single broad deuterium peak that implies the presence of a wide angular distribution in the orientational order, similar to the phenomenon observed in smectics with a high density of quenched disorder.

By measuring the self-diffusion coefficient of the water H$_2$O molecule in deuterated SDS samples, we were able to obtain the functional dependence of the relative self diffusion coefficient of the water molecules H$_2$O with respect to the total surfactant volume fraction (Fig. 3c). Often one can assume a single monolayer of surface-associated water. In this system however, there is the equivalent of approximately 3 to 4 layers of surface-associated water. This enhancement of water structure is interesting, and has indeed been reported before in high-salt conditions.

Diffusion time dependence on diffusion coefficients (Fig. 4a), linearity of the mean square displacement MSD versus time (but with non-zero offset) (Fig. 5a) and the contrast-matched diffusion experiments all point to a model that includes two ingredients: surfactant restricted diffusion in a cylindrical micelle, and micellar diffusion in water. Therefore, extracting the micellar diffusion from the slopes of the MSD curves (Fig. 5a) and fitting to a simple model with these two ingredients yields an average end-to-end distance of the wormlike micelle in the 1–1.5 µm range and is not strongly concentration-dependent (Fig. 5c), which is not far from the contour length found by Lopez-Diaz et al.

Rheology reports on longer timescales and lengthscales than NMR. There appears to be a distinct second relaxation time in the stress relaxation measurements, which is also consistent with oscillatory shear measurements (inset in Fig. 6a) that are not purely Maxwellian. This is consistent with previously reported results which showed deviations from the Maxwellian model at $R = 0.45$.

Finally, two distinct (Zimm and Rouse) polymer-like scalings are observed via NMR and rheometry respectively, indicating that the worm-like micelles exhibit classic polymer-like behaviour in different hydrodynamic regimes.

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**References**

Addition and correction

Note from RSC Publishing

This article was originally published with incorrect page numbers. This is the corrected, final version.

The Royal Society of Chemistry apologises for these errors and any consequent inconvenience to authors and readers.